Flow CAST®

Intended Use

The Flow CAST® is a Research Use Only test for the assessment of basophil activation upon stimulation with specific allergens. The test employs flow cytometry to determine the percent of basophil activation by measuring the number of basophils expressing the CD63 cell surface marker in K-EDTA whole blood samples.

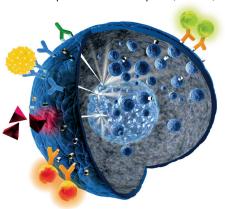
For laboratory use only.

Principle of the Assay

Flow CAST® is a basophil activation test (BAT) that reproduces an allergic reaction in a blood test. At the core of the assay is the ex vivo stimulation of basophils in an individual's whole blood with specific allergen, followed by flow cytometric determination of basophil activation.

Basophils are stained using two fluorescent labelled monoclonal antibodies: anti-CCR3-PE for selection and anti-CD63-FITC for the determination of the activation status. Basophils are identified by gating as CCR3pos/SSClow, and their activation status is assessed through CD63 expression.

The assay's readout is indicated as the ratio of CD63 positive basophils to total basophils (%CD63).



Interpretation of Results

Result	Interpretation
< cut-off	negative
≥ cut-off for one or both dilution of the allergen	positive

Cut-off

- Technical cut-off is 5% as established by reference range study
- Allergen-specific cut-off with improved specificity (Per CAST® Allergen List)

Clinical Study Performance

Allergen Group	Number of studies	Sensitivi- ty medi- an (range in %)	Allergic subjects (total)	Specifici- ty medi- an (range in %)	Control subjects (total)
Food Inhalants	5	92 (81-100)	311	93 (80-100)	240
Insect venoms	2	87 (73-89)	79	96 (95-97)	39
Drugs	7	55 (0-68)	227	91 (79-100)	167

11 peer-reviewed studies confirmed effectiveness in distinguishing between individuals with allergic disorders and non-allergic individuals. Allergic conditions were verified through individual's clinical history, oral food challenges, laboratory tests (such as skin prick tests and slgE), or a combination of these methods.

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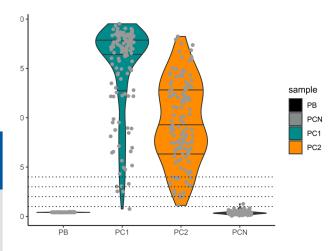
Performance Characteristics

Basophil Recovery

> 500 basophils/tube

Within-laboratory precision < 25 % CV for stimulus Performed according to CLSI guideline EP05-A3 and ISO standard 15197:2013

Reproducibility < 25 % CV for stimulus Performed according to CLSI guideline EP05-A3 and ISO standard 15197:2013.



Rate of IgE non-responder below 5%

In a study of 130 healthy blood donors, only 1 person had a poor response to Fc&RI receptor crosslinking, resulting in a non-responder rate of 0.77%. Most donors (81.5%) showed a strong response with an activation rate exceeding 50%.

Numbers of samples / per kit

Number of allergens	Individuals
1 / 1 conc.	25
2 / 1 conc.	20
3 / 1 conc.	16

Flow CAST®

Pre-Analytics

Sample required

EDTA whole blood

Sample storage

Store at 2 – 8°C (do not freeze)

Process within 48h

Drug allergens: Process within 24h

Collection

 K-EDTA venipuncture tubes must be filled at least half-way

No interference from the most common antihistamine were detected up to a specified concentration. For further detail refer to the IFU.

Processed Specimen Storage

Processed cells using standard protocol are fixed. Fixed cells may be stored at 2-8°C for 5 days for subsequent acquisition.

Equipment

- Flow cytometer equipped with a 488 nm (blue) laser source with emission filters for PE and FITC channel
- Water bath or incubator (37°C)
- Centrifuge
- Polystyrene or polypropylene tubes
- · Sterile or ultrapure water

CAST® Allergens

*ordered separately, refer to allergen list

Preparation:

- Add 250 µl Stimulation Buffer to prepared allergens
- Use freshly reconstituted allergens for stimulation
- Use up 4 stimulations for each allergen vial

Sample Preparation Layout

For each sample prepare single tubes for controls and desired allergens under examination.

PB = Sample Background

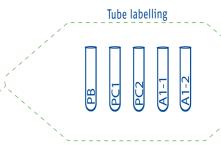
PC1 = Stimulation control with anti-Fc ERI AbFc

PC2 = Stimulation control with fMLP

A1-1 = Allergen 1 with dilution 1

A1-2 = Allergen 1 with dilution 2 etc.

Procedure (~ 60 minutes)



Stimulation and staining

Pipet to each tube:

- 1. 50 ul stimulus*
- 2. 100 ul stimulation buffer
- 3. 50 ul whole blood
- 4. 20 ul staining

Mix gently and incubate

*stimulus is not added to PB

Lysing of Red blood cells

Add 2.0 ml pre-warmed (18 - 28C) lysing reagent to all tubes



Incubate 5 - 10 min at 18 - 28C and centrifuge for 5 min at 500 x g

Add 300 ul* wash buffer containing fixative to resuspend the cell pellet

*to be adapted according to the Flow Cytometer in use

Flow Cytometry acquisition



Acquire the samples on the Flow cytometer

Fixation step: 30 min incubation (light protected) Storage: 2- 8 C for subsequent acquisition Acquire up to 5 days

Data Acquisition and Interpretation

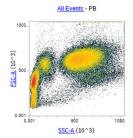


Figure 1: Three discrete populations (lymphocytes, monocytes and granulocytes) on an FSC/SSC

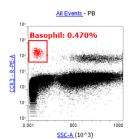


Figure 2: Selection of basophilic cells CCR3pos/SSClow

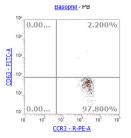


Figure 3: Sample Background (PB) with STB only

		Basophil - PC				
	104	0.00		95	5.455	0/0
	101				N-'	
CD63 - FITC-A	104			N. S.		
CD63 -	10)		\vdash	11.		_
	10-					
	10		103	10*	1.545	10
		9	CCR3 -	R-PE-A	7	

Figure 4: Stimulation Control (STCON)

Gated region	Count (n=)	
Total	125'864	100.0
Basophil	591	0.47
Q2 (CD63 ^{pos})	13	2.2
Q4 (CD63 ^{neg})	578	97.8

Gated region	Count (n=)	%
Total	130'926	100.0
Basophil	506	0.386
Q2 (CD63 ^{pos})	483	95.5
Q4 (CD63 ^{neg})	23	4.5

This document is for information purpose only, before performing the assay please carefully refer/read the respective IFU available (https://www.buhlmannlabs..com/cellu-lar-alleray/flow-cast

Ordering code:

FK-CCR-U 100 tests

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BÜHLMANN Laboratories AG Switzerland info@buhlmannlabs.ch www.buhlmannlabs.ch Phone +41 61 487 12 12 BUHLMANN Diagnostic Corp USA info@buhlmannlabs.com www.buhlmannlabs.com Phone 844 300-9799