



BÜHLMANN GanglioCombi® MAG ELISA

with enzyme labels IgG/IgM Mix, IgG and IgM

Detection of anti-ganglioside
and -MAG antibodies by ELISA
(HNK-1 ("MAG"), GM1, GT1a GD1a, GD1b and GQ1b)

For research use only.
Not for use in diagnostic procedures.

EK-GCM-U 2 x 96 tests

Release Date: 2025-08-25
Version A2

ENGLISH

INTENDED USE

The BÜHLMANN GanglioCombi® MAG ELISA is an assay for the semi-quantitative determination of IgG and/or IgM antibodies against selected neural antigens/epitopes in serum samples.

For research use only. Not for use in diagnostic procedures.

INTENDED APPLICATION

The three different enzyme labels, provided in the kit, enable three different testing algorithms:

1. Testing with the IgG/IgM conjugate mix (hereafter referred to as mix) allows to screen for the presence of anti-neutral antibodies.
2. Testing with individual IgG and/or IgM conjugates allows antibody isotype determination.
3. For laboratory work-up initial sample screening using the mix (option 1), may be followed by differentiation of mix-positive samples using individual IgG and IgM conjugates (option 2), if required.

PRINCIPLE OF THE ASSAY

The BÜHLMANN GanglioCombi® MAG ELISA allows the measurement of ganglioside and Myelin Associated Glycoprotein (MAG) antibodies in serum. The microtiter plate is coated with gangliosides: GM1, GT1a, GD1a, GD1b, GQ1b and the chemically synthesized HNK-1 epitope of the MAG glycoprotein.

Serum samples, controls and calibrator are added to wells of the microtiter plate. After 2 hours of incubation at 2-8°C and washing steps, detection antibodies (anti-IgG/IgM, anti-IgG, anti-IgM) conjugated to horseradish peroxidase (HRP) detect the anti-ganglioside and/or anti-MAG antibodies bound to the immobilized gangliosides or HNK-1 on the plate. After another 2 hours of incubation and further washing steps, the chromogenic HRP substrate, tetramethylbenzidine (TMB) is added (blue color formation) followed by a stopping reaction (change to yellow color). The absorption is measured at 450 nm.

The measured absorbance is proportional to the titer of antibodies present in a given sample. Antibody titers are expressed as % Ratios of the calibrator.

REAGENTS SUPPLIED AND PREPARATION

Reagents	Quantity	Code	Reconstitution
Microtiter Plate precoated with gangliosides and HNK-1	2 x 12 x 8 wells strips with frame	B-GCM-MP	Ready to use
Plate Sealer	6 pieces		
Wash Buffer Concentrate (10x) with preservatives	2 bottles 100 mL	B-GCO-WB	Dilute with 900 mL of deionized water
Incubation Buffer with preservatives	1 bottle 100 mL	B-GCO-IB	Ready to use
Calibrator Lyophilized with preservatives	1 vial	B-GCO-CA	Add 1.5 mL of Incubation Buffer
Control Negative, Low and Medium ¹ Lyophilized with preservatives	3 vials	B-GCO-CONSET	Add 1.5 mL of Incubation Buffer
Enzyme Label IgG/IgM Mix Anti-human IgG and IgM antibody conjugated to HRP in a buffer matrix with preservatives	2 vials 11 mL each	B-GCO-ELGM	Ready to use
Enzyme Label IgG Anti-human IgG antibody conjugated to HRP in a buffer matrix with preservatives	1 vial 11 mL	B-GCO-ELG	Ready to use
Enzyme Label IgM Anti-human IgM antibody conjugated to HRP in a buffer matrix with preservatives	1 vial 11 mL	B-GCO-ELM	Ready to use
TMB Substrate TMB in citrate buffer	2 vials 11 mL	B-TMB	Ready to use
Stop Solution 0.25 M sulfuric acid	2 vials 11 mL	B-STTS	Ready to use Corrosive agent

Table 1

¹ The controls contain lot specific levels of anti-GM1 antibodies. Refer to the additional QC data sheet for actual mean OD and % Ratio.

STORAGE AND SHELF LIFE OF REAGENTS

Sealed/ unopened reagents	
Store at 2-8 °C. Do not use the reagents beyond the expiration date printed on the labels.	
Opened/ reconstituted reagents	
Microtiter Plate	Return unused strips immediately to the foil pouch containing the desiccant packs and reseal along the entire edge of zip-seal. Store for up to 6 months at 2-8 °C.
Diluted Wash Buffer	Store for up to 6 months at 2-8 °C.
Incubation Buffer	
Enzyme Labels	
TMB Substrate	
Calibrator	
Controls	
Stop Solution	Store for up to 6 months at 18-28 °C.

Table 2

MATERIALS REQUIRED BUT NOT PROVIDED

- Precision pipettes with disposable tips: 10 µL, 20 µL, 100 µL and 1000 µL pipettes
- Disposable polystyrene or polypropylene tubes for the preparation of sample dilutions
- 1000 mL cylinder for the dilution of the wash buffer
- Microtiter plate washer
- Blotting paper
- Microtiter plate shaker
- Microtiter plate reader for the measurement of absorbance at 450 nm

WARNINGS AND PRECAUTIONS

Safety precautions

- The calibrator and controls of this kit contain components of human origin. Although tested and found negative for HBV, HCV and HIV1/2, the reagents should be handled as if capable of transmitting infections and should be handled in accordance with Good Laboratory Practices (GLP) using appropriate precautions.
- This kit contains components classified in accordance with the Regulation (EC) No. 1272/2008:
 - The stop solution contains sulfuric acid (conc. 2.5 – 5%), thus the reagents may cause skin irritation (H315), serious eye irritation (H319), and may be corrosive to metals (H290).
 - The calibrator, controls and enzyme labels contain 2-methyl-4-isothiazolin-3-one hydrochloride (conc. $\geq 0.0015\%$), thus the reagents may cause allergic skin reactions (H317).
 - The incubation buffer and wash buffer contain gentamicin sulphate, thus, the reagents may cause an allergic skin reaction (H317).
- Avoid contact of reagents with the skin, eyes, or mucous membranes. If contact does occur, immediately wash with generous amounts of water; otherwise, irritation / burns can occur.
- Reagents and chemicals have to be treated as hazardous waste according to the national biohazard safety guideline or regulation.

Technical precautions

- Read the instructions carefully prior to carrying out the test. Test performance will be adversely affected, if reagents are incorrectly diluted, modified or stored under conditions other than those as detailed in this instruction for use.

ELISA procedure

Temperature of reagents

- Prepare reagents before starting the assay procedure. Steps 3-9: Reagents used in steps 3-9 must be cold (2-8 °C) and kept cold while pipetting and washing. Recommendation: Prepare the wash buffer the day before performing the assay and place it into the fridge overnight.
- Perform all wash steps with cold (2-8 °C) wash buffer.

- Adjust TMB substrate and stop solution to room temperature (18-28 °C) at the start of the assay procedure.

Washing steps

- Wash steps 3, 6 and 9 are crucial to remove residues resulted from the production process and/or potentially unbound antibodies in the wells.
- An automated washer operating in “plate mode” is strongly recommended, i.e. each process step (dispense / aspiration) is carried out on all of the strips, sequentially, before the instrument continues with the next washing cycle.

Substrat incubation

- Step 11: Shake the microtiter plates during incubation with substrate. Depending on the model of the plate shaker we recommend 400-600 rpm. The solution should move in the wells but must not spill over.

Kit components

- Components must not be used after the expiry date printed on the labels.
- Do not mix different lots of reagents.
- Every effort should be made to ensure that no cross contamination occurs between reagents, samples or between wells.
- Microwells cannot be re-used.

SPECIMEN COLLECTION AND STORAGE

The procedure requires <0.1 mL of blood or <50 µL of serum, respectively.

Collect blood into plain venipuncture tubes without any additives and avoid hemolysis. Perform serum preparation according to manufacturer's instructions. Decant the serum.

Serum samples can be stored at 2-8 °C for up to eight weeks, at 28 °C for up to one week and at ≤ -20 °C for 16 weeks. Frozen samples should be thawed and mixed thoroughly by gentle swirling or inversion prior to use. We recommend preparing aliquots of serum samples before freezing in order to avoid repeated freeze/thaw cycles.

ASSAY PROCEDURE

There are two options:

- (1) Detection of mix-isotypes (IgG and IgM): add enzyme label mix in step 7
- (2) Detection of IgG or IgM isotypes: add enzyme label IgG or enzyme label IgM in step 7

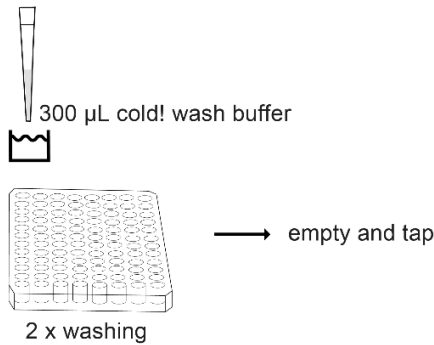
Note: Adjust TMB substrate solution to room temperature (18-28 °C).

1. Dilute samples 1:50 with incubation buffer. Use e.g. 20 µL of serum + 980 µL of cold! (2-8 °C) incubation buffer. Mix thoroughly by vortexing and leave diluted samples as well as reconstituted calibrator and controls at 2-8 °C for 30 minutes prior to pipetting (refer to step 4a and b).

2. Prepare a plate-frame with sufficient strips to test the required number of calibrators, controls, and samples. Remove excess strips from the frame and reseal it in the foil pouch together with the desiccant packs without delay. Store refrigerated.

Note: Use cold reagents in steps 3 to 9.

3. Wash the wells twice using at least 300 μL of cold! (2-8 $^{\circ}\text{C}$) wash buffer per well. Empty the wells and tap the plate firmly onto blotting paper to remove remaining liquid completely.



Note: Immediately proceed to the next steps.

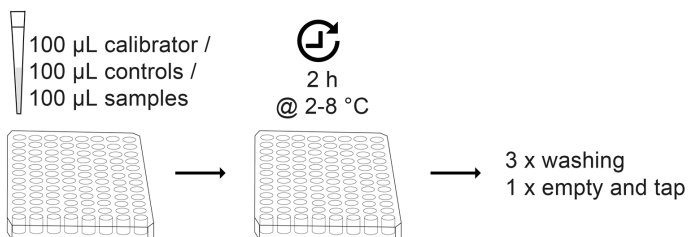
- 4a. Pipet 100 μL of calibrator into the well A1 (refer to figure 1A or figure 1B for option 2).
- 4b. Pipet 100 μL of medium control into well B1, of low control into well A2 and of negative control into well B2 (refer to figure 1A or 1B).

Note for option 1: If more than three strips per run are used, calibrator and controls can be tested in duplicates (see figure 1A).

- 4c. Pipet 100 μL of diluted sample 1 into wells C1-H1 (refer to figure 1A or 1B).
- 4d. Pipet 100 μL of diluted sample 2 into wells C2-H2 (refer to figure 1A or 1B).
- 4e. Pipet 100 μL of diluted samples 3-24 (for option 1) or 3-12 (for option 2) into subsequent wells (refer to figure 1A or 1B).

Note for option 2: Repeat the pipetting of samples 1-12 in the same order into the remaining wells for testing with the second isotype.

5. Cover the plate with a plate sealer and incubate for 2 hours (± 5 min) at 2-8 $^{\circ}\text{C}$ (do not shake the plate).
6. Remove the plate sealer. Empty the wells and wash three times using at least 300 μL of cold! (2-8 $^{\circ}\text{C}$) wash buffer per well. Empty the wells and tap the plate firmly onto blotting paper in order to remove wash buffer completely.

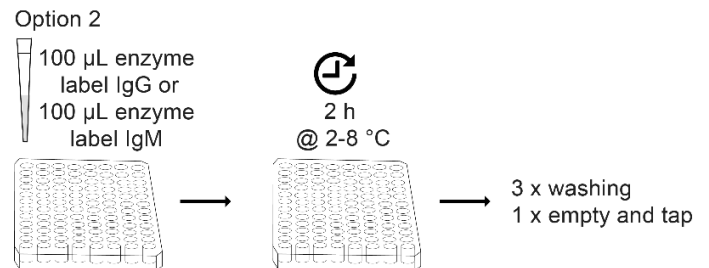
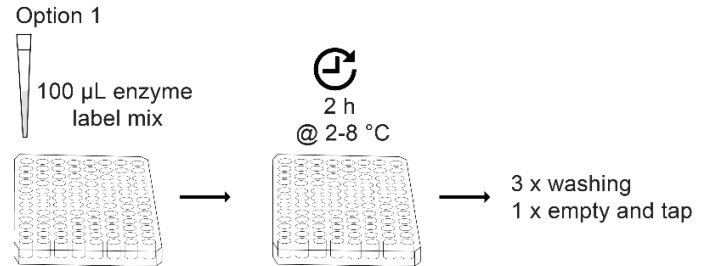


For option 1: Detection of mix-isotype

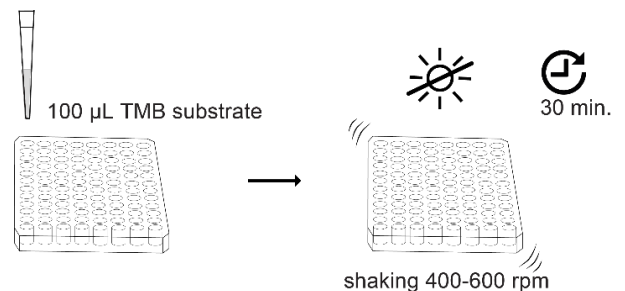
7. Add 100 μL of mix to the wells.

For option 2: Detection of IgG or IgM isotypes

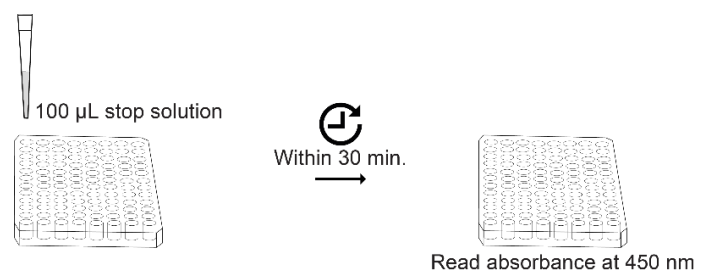
- 7'. Add 100 μL of either enzyme label IgG or IgM to the respective wells (refer to figure 1B).
8. Cover the plate with a plate sealer and incubate for 2 hours (± 5 min) at 2-8 $^{\circ}\text{C}$ (do not shake the plate).
9. Remove the plate sealer. Empty the wells and wash three times using at least 300 μL of cold! (2-8 $^{\circ}\text{C}$) wash buffer per well. Empty the wells and tape the plate firmly onto blotting paper.



10. Add 100 μL TMB substrate solution (equilibrated to room temperature) to each well.
11. Cover the plate with a plate sealer, protect the plate from light and incubate on a plate shaker set at 400-600 rpm, at 18-28 $^{\circ}\text{C}$ for 30 ± 2 minutes.



12. Add 100 μL of stop solution to all wells. Remove air bubbles with a pipette tip. Proceed to step 13 within 30 minutes.
13. Read the absorbance at 450 nm in a microtiter plate reader.



QUALITY CONTROL

Thorough understanding of this instruction for use is necessary for the successful use of the product. Reliable results will be obtained only by using precise laboratory techniques and accurately following this instruction for use. The BÜHLMANN GanglioCombi® MAG ELISA kit comes with three controls: negative, low and medium control. The controls have assigned value ranges (% Ratio) indicated on the QC-data sheet supplied with each kit. The control measurements must be within the indicated value ranges to obtain valid results. In addition to kit controls, we recommend the use of serum pools for internal quality control.

A minimal OD_{450nm} value of 1.2 is recommended for the calibrator.

Performance characteristics should be within established limits. If the performance of the assay does not meet the established limits and repetition has excluded errors in technique, check the following issues: i) temperature controlling (reagents used in step 3-9 kept at 2-8 °C); ii) accuracy of thermometers, pipetting and timing devices; iii) ELISA reader settings; iv) expiration dates of the reagents v) storage and incubation conditions; vi) color of the TMB substrate solution (should be colorless); vii) purity of water; and viii) aspiration and washing methods.

STANDARDIZATION AND METROLOGICAL TRACEABILITY

There are no internationally or nationally recognized reference materials or reference measurement procedures for anti-ganglioside or -MAG antibodies in serum samples. The BÜHLMANN GanglioCombi® MAG ELISA is standardized against an internally established reference material. Calibrator values are assigned according to a value transfer protocol (ref. 1), to guarantee metrological traceability, and are indicated in arbitrary "% Ratio" units.

The 95% confidence interval of the combined uncertainty of product calibrators was determined to be 29.3% for IgG antibodies and 37.6% for IgM antibodies.

CALCULATION OF TEST RESULTS

1. Record absorbance (OD) at 450 nm for each well (calibrator, controls and samples).
2. If multiple calibrator and control measurements were performed, average the values.

Results are expressed as Ratio of absorbance of samples and the (averaged) absorbance of the calibrator.

IgG/IgM Mix isotypes

$$\% \text{ Ratio} = \frac{\text{absorbance of samples or controls}}{\text{absorbance of calibrator}} \times 200$$

IgG and IgM isotypes

$$\% \text{ Ratio} = \frac{\text{absorbance of samples or controls}}{\text{absorbance of calibrator}} \times 100$$

Programs to calculate results as % Ratio are available on most microplate readers.

Note: Results presented in tables 3 and 4 are examples and are provided for demonstration purpose only.

LIMITATIONS

- This test has not been validated for plasmapheresis samples.
- Intravenous immunoglobulins (IVIg) may affect test results.

PERFORMANCE CHARACTERISTICS

Method comparison

BÜHLMANN GanglioCombi® MAG ELISA vs anti-MAG Antibodies ELISA

The method comparison study was performed according to the CLSI guideline EP09-A3 and EP12-A2. One hundred and twenty-two (122) samples were measured using 2 lots of BÜHLMANN GanglioCombi® MAG ELISA and 2 lots of anti-MAG Antibodies ELISA. Diagnostic (kappa) agreement, negative percent agreement and positive percent agreement were determined. The agreements are presented in table 5.

Within-laboratory precision

For anti-gangliosides: 5.7 – 13.2% CV

For anti-MAG: 14.4 – 36.5% CV

Within-laboratory precision was established according to the CLSI guideline EP05-A3 using the standardized 20 days x 2 runs x 2 replicates study design. Three (3) pooled serum samples were tested. The results are summarized in table 6.

Reproducibility

For anti-gangliosides: 7.7 – 19.1% CV

For anti-MAG: 23.5 – 33.2% CV

Reproducibility was established according to the CLSI guideline EP05-A3 using a 3 instrument/lot/operator x 5 days x 5 replicates study design. Three (3) pooled serum samples were tested. The results are summarized in table 7.

Limit of blank (LoB) ≤ Limit of detection (LoD): ≤30% Ratio

The LoB and LoD was established according to the CLSI guideline EP17-A using the non-parametric analysis. The results are summarized in table 8.

High dose hook effect

No limitation due to a high dose hook effect to the measuring range was observed.

INTERFERING SUBSTANCES

The susceptibility of the assay to oral and injectable pharmaceuticals, as well as to endogenous substances was assessed according to CLSI guideline EP07-A3. Bias in results $\geq \pm 20\%$ Ratio was considered interference.

No interference is detected with the following substances up to the listed concentrations: intravenous immunoglobulin (20 mg/mL), rituximab (3 mg/mL), cladribine (273 ng/mL), Interferon alpha-2a (49.5 ng/mL), gabapentin (26.7 µg/mL), ibuprofen (0.22 mg/mL), chlorambucil (1.96 µg/mL), prednisone (99 ng/mL), prednisolone (1.2 µg/mL), rheumatoid factor (2340 IU/mL), hemoglobin (10 mg/mL), hemolysate (10 mg/mL), triglyceride (15 mg/mL), conjugated bilirubin (20 µg/mL), unconjugated bilirubin (150 µg/mL).

TABLES AND FIGURES

Microtiter plate set-up: IgG/IgM-Mix label

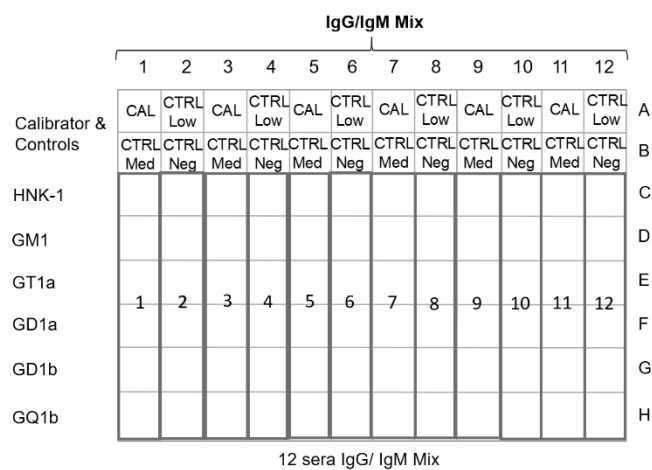


Figure 1A: ≤ 24 sera / Kit (2 MP / Kit)

Microtiter plate set-up: IgG & IgM labels

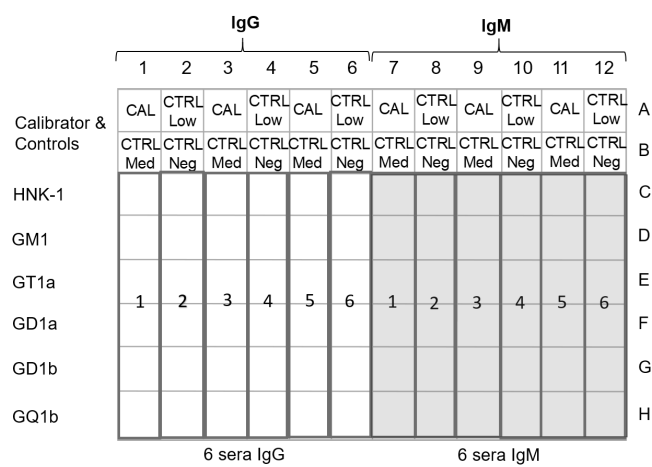


Figure 1B: 2 profiles / serum, ≤ 12 sera / Kit (2 MP / Kit)

Example of results

A IgG/IgM-Mix label

B-GCO-ELGM	Absorbance (OD450)	Ratio [%]
Calibrator	2.250	
	2.276	
Calibrator Avg.	2.263	200
Medium Control	1.834	
	1.880	
Medium Control Avg.	1.857	164
Low Control	0.513	
	0.510	
Low Control Avg.	0.512	45
Negative Control	0.040	
	0.041	
Negative Control Avg.	0.041	4
Sample 1 HNK-1	0.299	26
Sample 1 GM1	0.264	23
Sample 1 GT1a	0.500	44
Sample 1 GD1a	0.200	18
Sample 1 GD1b	1.018	90
Sample 1 GQ1b	0.228	20

Table 3

B IgG & IgM labels

Enzyme label	Absorbance (OD450)		Ratio [%]	
B-GCO-ELG/ B-GCO-ELM	IgG	IgM	IgG	IgM
Calibrator	2.488	2.411		
	2.446	2.201		
Calibrator Avg.	2.467	2.306	100	100
Medium Control	1.879	1.734		
	1.987	1.818		
Medium Control Avg.	1.933	1.776	78	77
Low Control	0.452	0.501		
	0.716	0.609		
Low Control Avg.	0.584	0.555	24	24
Negative Control	0.045	0.048		
	0.037	0.042		
Negative Control. Avg.	0.041	0.045	2	2
Sample 1 HNK-1	0.423	0.621	17	27
Sample 1 GM1	2.001	2.102	81	91
Sample 1 GT1a	0.521	0.237	21	10
Sample 1 GD1a	1.984	0.821	80	36
Sample 1 GD1b	0.473	1.923	19	83
Sample 1 GQ1b	0.094	0.911	4	40

Table 4

TABLES AND FIGURES

Method comparison anti-MAG antibodies

Description	N	Kappa agreement		NPA		PPA	
		Value	95% CI	Value	95% CI	Value	95% CI
EK-GCM IgM vs. EK-MAG	122	0.88	0.80 - 0.97	100.0%	94.6% - 100.0%	87.5%	75.9% - 94.8%
EK-GCM IgG/IgM Mix vs. EK-MAG	122	0.87	0.78 - 0.96	97.0%	89.5% - 99.6%	89.3%	78.1% - 96.0%

Table 5

NPA: Negative Percent Agreement
PPA: Positive Percent Agreement
CI: Confidence Interval

Within-laboratory precision

Sample Description		Within-Laboratory Precision			
Analyte	Enzyme Label (Isotype)	N	Mean [%Ratio]	SD [%Ratio]	CV [%]
anti-GM1 Ab	IgM	80	48	3.5	7.2
		80	91	6.2	6.8
	IgG	80	40	5.1	12.9
		80	106	13.1	12.4
anti-GQ1b Ab	IgM	80	45	2.6	5.7
		80	85	6.7	7.8
	IgG	80	43	5.7	13.2
		80	80	6.9	8.6
anti-MAG Ab	IgM	80	34	6.3	18.7
		80	72	10.4	14.4
	IgGM	80	27	9.6	35.3
		80	51	18.8	36.5

Table 6

Reproducibility

Sample Description		Reproducibility			
Analyte	Enzyme Label (Isotype)	N	Mean [%Ratio]	SD [%Ratio]	CV [%]
anti-GM1 Ab	IgM	75	51	4.9	9.7
		75	94	7.2	7.7
	IgG	75	39	5.6	14.5
		75	106	17.1	16.1
anti-GQ1b Ab	IgM	75	48	3.9	8.2
		75	92	9.9	10.7
	IgG	75	42	8.1	19.1
		75	78	12.0	15.4
anti-MAG Ab	IgM	75	43	14.3	33.2
		75	98	23.1	23.5
	IgGM	75	42	10.6	25.0
		75	97	27.2	28.0

Table 7

LoD and LoB

Analyte	LoB [% Ratio]	LoD [% Ratio]
Anti-GM1 IgM Ab	5	21
Anti-GM1 IgG Ab	6	15
Anti-MAG IgM Ab	12	26
Anti-MAG IgG/IgM Mix Ab	14	27
Anti-GQ1b IgM Ab	3	17
Anti-GQ1b IgG Ab	8	18

Table 8

SHORT PROTOCOL

Important: The short protocol is not a substitute for the detailed information described in this instruction for use.

Before testing day

Wash Buffer Preparation

Dilute wash buffer concentrate
1:10 with deionized water



Recommendation: Prepare the wash buffer the day before performing the assay and place it into the fridge overnight.

Testing day

Samples / Controls / Calibrator Preparation

Dilute serum samples 1:50 with
(cold)! incubation buffer and mix
thoroughly by vortexing

Reconstitute controls and
calibrators



leave 30 minutes at 2-8 °C

BÜHLMANN GanglioCombi® MAG ELISA

Precoated microtiter plate



wash 2 x with $\geq 300 \mu\text{L}$ (cold)! wash buffer

100 μL calibrator, controls or
serum samples (1:50)



incubate 2 hours (± 5 min) at 2-8 °C

wash 3 x with $\geq 300 \mu\text{L}$ (cold)! wash buffer

add 100 μL enzyme label(s)



incubate 2 hours ± 5 min at 2-8 °C

wash 3 x with $\geq 300 \mu\text{L}$ (cold)! wash buffer

add 100 μL TMB substrate (ambient temperature)!



*incubate 30 min (± 2 min) at 18-28 °C
on a plate rotator*

add 100 μL stop solution (ambient temperature)!

➡ Read absorbance at 450 nm (within 30 minutes)

TIME TO RESULT: 5 HOURS

REFERENCES

1. Blirup-Jensen, S., Johnson, A. M. & Larsen, M. Protein standardization V: Value transfer. A practical protocol for the assignment of serum protein values from a Reference Material to a Target Material. *Clin. Chem. Lab. Med.* **46**, 1470–1479 (2008).

CHANGELOG

Date	Version	Change
2025-08-25	A2	Revision of chapter <i>Tables and figures (Table 3)</i> , <i>Short protocol</i> and <i>Symbols</i>

SHIPPING DAMAGE

Please notify your distributor, if this product was received damaged.

SYMBOLS

BÜHLMANN use symbols and signs listed and described in ISO 15223-1.

For definition of symbols see the symbol glossary at: www.buhlmannlabs.ch/support/downloads/

In addition, the following symbols and signs are used:

Symbol	Explanation
MP	Microtiter Plate
BUF WASH 10X	Wash Buffer concentrate (10x)
BUF INC	Incubation Buffer
CAL	Calibrator
CONTROL -	Control Negative
CONTROL L	Control Low
CONTROL M	Control Medium
EL IgG	Enzyme Label IgG
EL IgM	Enzyme Label IgM
EL MIX	Enzyme Label IgG/IgM Mix
SUBS TMB	TMB Substrate
SOLN STOP	Stop Solution